



## Nephroprotective Effect of a Combination of Probiotics, Tender Coconut Water, and Vitamin E in Gentamicin-Induced Acute Kidney Injury in Rats

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### ARTICLE INFO

#### Article history:

Received 22 March 2025

Revised 24 May 2025

Accepted 02 June 2025

Published online 01 August 2025

### ABSTRACT

Gentamicin increases Reactive Oxygen Species (ROS), leading to cell damage and kidney necrosis. The body counteracts free radicals by producing endogenous antioxidants. Natural antioxidants such as probiotics, vitamin E, and phenolic compounds are known to protect the kidneys. This study aims to evaluate the nephroprotective effect of a combination of probiotics, tender coconut water, and vitamin E on endogenous antioxidants in rats with an Acute Kidney Injury (AKI) model induced by gentamicin. The parameters measured were superoxide dismutase (SOD) and glutathione peroxidase (GPx). The study used a posttest-only control group design with 30 male Wistar rats divided into five groups: K1 (normal); K2 (gentamicin-induced); K3 (gentamicin + tender coconut water 8 mL/200 g BW); K4 (gentamicin + tender coconut water + probiotics); and K5 (gentamicin + tender coconut water + probiotics + vitamin E 1.8 IU/200 g BW). Gentamicin was administered at 20 mg/200 g BW/day via intraperitoneal injection. Data were analyzed using One-way ANOVA. The mean SOD levels in K2 were  $23.77 \pm 3.07\%$ , increasing to  $66.39 \pm 3.07\%$  in K5. The mean GPx levels in K2 were  $23.77 \pm 0.75$  u/mg, rising to  $61.94 \pm 0.72$  u/mg in K5. SOD and GPx levels were significantly higher in K5 compared to K2. The combination of probiotics, tender coconut water, and vitamin E exerts a nephroprotective effect in AKI model rats induced by gentamicin, as indicated by increased SOD and GPx levels compared to the negative control.

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**Keywords:** Probiotics, tender coconut water, vitamin E, nephroprotective, endogenous antioxidants.

### Introduction

Gentamicin is an aminoglycoside antibiotic used for infections caused by gram-negative bacteria. In addition to being useful as an antibiotic, gentamicin also has side effects if given in excessive doses, namely damage to the kidneys (nephrotoxic).<sup>1</sup> Kidney damage due to gentamicin in high doses will accumulate in the proximal tubule after being filtered in the glomerulus.<sup>2</sup> Accumulation of gentamicin in cells can cause an increase in reactive oxygen species (ROS), which can then trigger an increase in free radicals, causing cell damage and kidney necrosis.<sup>3</sup> Gentamicin can reduce endogenous antioxidants and increase ROS production, which can trigger oxidative stress. Gentamicin increases the formation of superoxide anions, hydrogen peroxide, and hydroxyl radicals and releases iron (Fe) from the mitochondria of the renal cortex, which causes lipid peroxidation *in vitro* and is a strong catalyst for the formation of free radicals.<sup>4</sup> Nephrotoxicity caused by gentamicin can be through oxidative stress with excessive ROS production.<sup>5</sup> The body's response to counteract free radicals is to produce endogenous antioxidants such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx). Still, if the free radicals formed exceed endogenous antioxidants, exogenous antioxidants are needed to create a balance. Exogenous antioxidants from natural sources obtained from plants are essential.<sup>6</sup>

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**Citation:** Zulaikhah ST, Ratnawati R, Hussaana A. Nephroprotective Effect of a Combination of Probiotics, Tender Coconut Water, and Vitamin E in Gentamicin-Induced Acute Kidney Injury in Rats. Trop J Nat Prod Res. 2025; 9(7) 3176 – 3180 <https://doi.org/10.26538/tjnpr/v9i7.42>

Official Journal of Natural Product Research Group, Faculty of Pharmacy, University of Benin, Benin City, Nigeria

Several natural exogenous antioxidants such as probiotics, vitamin E, and phenolic compounds found in tender coconut water are known to protect against nephrotoxicity (nephroprotective). Research conducted by Zulaikhah *et al.*<sup>7</sup> showed that coconut water has an antioxidant effect on kidney cell damage caused by increased free radicals triggered by exposure to lead. Kunle-Alabi *et al.*<sup>8</sup> reported that coconut water has a protective effect on the kidneys and liver caused by exposure to monosodium glutamate in mothers and their offspring. Ekezie *et al.*<sup>9</sup> reported that tender coconut water protects against tetrachloride-induced kidney damage in Wistar rats. Research conducted by Nwangwa<sup>10</sup> showed that giving coconut water can prevent kidney damage in a rat model of alloxan-induced diabetes mellitus. Chrisanto<sup>11</sup> reported that probiotic drinks from tender coconut water have a strong antioxidant effect. Vitamin E has an antioxidant effect by eradicating peroxyl radicals, regulating oxidative changes in cell organelles, preventing lipid peroxidation, and preventing the destruction of kidney cells.<sup>12</sup> Nugraha *et al.*<sup>13</sup> also reported that Vitamin E exerts a protective effect on kidney damage in white rats induced with ethinyl estradiol. This study aims to evaluate the nephroprotective effect of a combination of probiotics, tender coconut water, and Vitamin E on endogenous antioxidants in rat models of Acute Kidney Injury due to Gentamicin.

### Materials and Methods

#### Experimental animal

Ethical clearance for this study was obtained from the Medical/Health Research Bioethics Commission of the Faculty of Medicine, Unissula Semarang, with reference number 380/IX/2024/Komisi-Bioetik. The treatment of experimental animals and the examination of SOD and GPx, levels were conducted at the Center for Food & Nutrition Studies (PSPG) Gadjah Mada University Yogyakarta.

### Gentamicin induction

Gentamicin administration was carried out simultaneously with the administration of probiotics made from tender coconut water and vitamin E, with a dose of 100 mg/kg BW/day via Intraperitoneal injection.<sup>14</sup>

### Administration of tender coconut water

Tender coconut water from green coconuts around 5-7 months old was obtained from the Yogyakarta area and its surroundings. The tender coconut water was given to the treatment group at 8 mL/200 g BB/day two times and administered for 14 days. The dose was adjusted to previous research and given orally by sonde for 2 weeks.<sup>15</sup>

### Probiotic administration

Probiotics from product X containing *Lactobacillus* were added to tender coconut water at a dose of 18 mg/200 g BW for every 8 mL of tender coconut water, given orally using a tube for 2 weeks.<sup>15</sup>

### Administration of vitamin

The dose of vitamin E was 1.8 IU/200 g BW, given orally for 2 weeks.<sup>16</sup>

### Provision of treatment

Healthy Male Wistar rats (2 months old, 180-220 g) were used in this experiment. A total of 30 rats were randomly divided into 5 groups of 6 rats each.

### Blood collection

Blood was withdrawn by inserting the microhematocrit tube into the ophthalmic vein in the corner of the mouse's eyeball periorbital, then rotated slowly until blood flowed and collected in 2 mL of Eppendorf tubes.<sup>17</sup>

### Antioxidant assays

Examination of endogenous antioxidants by measuring SOD and GPx levels using the Elisa method.<sup>17</sup>

### Statistical analysis

All values are expressed as mean  $\pm$  SD. Data between groups were first tested by one-way ANOVA and then between two groups were analyzed by Pots Hoc Least Significant Different (LSD). Statistical analyses were performed using SPSS-26. A P value < 0.05 was considered statistically significant.<sup>18</sup>

## Results and Discussion

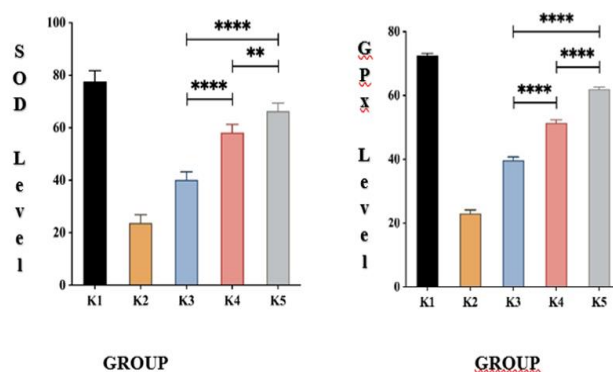
The effect of the combination of probiotics, tender coconut water, and vitamin E on endogenous antioxidants is presented in Table 2. Indicators of endogenous antioxidants measured in this study were glutathione peroxidase (GPx) and Superoxide dismutase (SOD) levels. Data presented in Table 2 shows that the results of the normality test with Shapiro Wilk and homogeneity with Levene test for SOD and GPx levels have values >0.05. This indicates that both variables have normal and homogeneous data distribution, so the data were analyzed using the One-way ANOVA test. The lowest average SOD level in K2 was 23.77%, and the highest level in K1 was 77.59%. The average SOD level in K3 was 40.16%, K4 was 58.20, and K5 was 66.39%. The results of the ANOVA test showed a p-value: 0.0001, meaning that there was a significant difference in SOD levels in the five groups. The average GPx level observed in K2 was 23.77 u/mg, and the highest level observed in K1 was 72.41 u/mg. The average level of GPx in K3 was 39.73 u/mg, K4 was 51.43 u/mg, and K5 was 61.94 u/mg. The results of the ANOVA test showed a p-value of 0.0001, meaning there was a significant difference in GPx levels in the five groups. The results of this analysis indicate that the combination of probiotics, tender coconut water, and Vitamin E enhanced the activity of the endogenous antioxidants in gentamicin-induced acute kidney injury. Hence, combining probiotics, tender coconut water, and Vitamin E exerted a nephroprotective effect.

**Table 1: Groups and treatments**

Group	Treatment <sup>a</sup>
Group 1 (K1):	The healthy rat group, namely male albino rats of the Wistar strain, received a standard feed diet and drinking ad libitum
Group 2 (K2):	The negative control group was male albino Wistar rats induced by gentamicin and given a standard feed diet and drinking ad libitum
Group 3 (K3):	Treatment group 1, namely male albino Wistar rats induced by gentamicin and given a standard feed diet, drinking <i>ad libitum</i> and being given tender coconut water at a dose of 8 mL/200 g BW for 2 weeks
Group 4 (K4):	Treatment group 2, namely male albino Wistar rats induced by gentamicin and given a standard feed diet, drinking <i>ad libitum</i> and being given tender coconut water at a dose of 8 mL/200 g BW mixed with probiotics at a dose of 18 mg for 2 weeks.
Group 5 (K4):	Treatment group 3, namely male albino Wistar rats induced by gentamicin and given a standard feed diet, drinking <i>ad libitum</i> , tender coconut water at a dose of 8 mL/200 g BW mixed with probiotics at a dose of 18 mg and vitamin E at a dose of 1.8 IU/200 g BW for 2 weeks. <sup>15</sup>

After the treatment is complete, blood is taken to measure endogenous antioxidants, namely SOD and GPx levels.

Analysis of Figure 1 shows that the highest average levels of both SOD and GPx were seen in the group of mice induced by gentamicin and given tender coconut water with probiotics and vitamin E. This was also observed in the the group of mice induced by gentamicin and given tender coconut water with probiotics and vitamin E.



**Figure 1:** Results of the analysis of differences in mean SOD and GPx levels between groups using the LSD Post Hoc Test.

**Table 2:** Average, normality test, homogeneity test, and anova test for SOD and GPx levels in 5 groups

Variable (oxidative stress)	Group					p value
	K1 <sup>a</sup> Average ± SD	K2 <sup>b</sup> Average ± SD	K3 <sup>c</sup> Average ± SD	K4 <sup>d</sup> Average ± SD	K5 <sup>e</sup> Average ± SD	
Levels of SOD (%)	77.59 ± 4.23	23.77 ± 3.07	40.16 ± 3.07	58.20 ± 3.07	66.39 ± 3.07	
Shapiro-Wilk Test	0.964	0.961	0.910	0.961	0.960	>0.05*
Levene's Test						>0.05*
One way ANOVA						0.0001**
Level of GPx (u/mg)	72.41 ± 0.75	23.77 ± 0.75	39.73 ± 1.02	51.43 ± 0.92	61.94 ± 0.72	
Shapiro-Wilk Test	0.796	0.459	0.649	0.849	0.976	>0.05*
Levene's Test						>0.05*
One way ANOVA						0.0001**

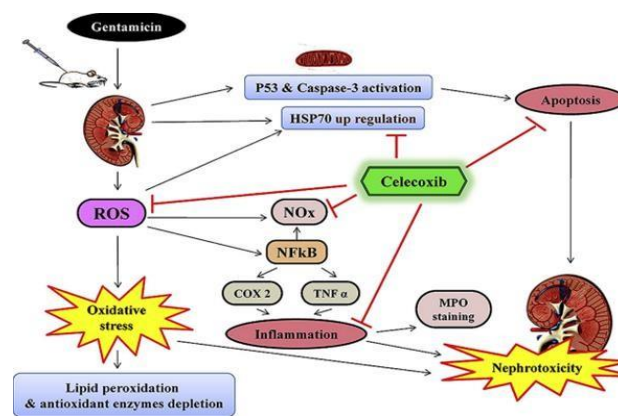
\*: Significant

a: Healthy rat group, namely male white rats of the Wistar strain that received a standard feed diet and drink ed libitum, b: Negative control group, namely male white rats of the Wistar strain that were induced by gentamicin and received a standard feed diet and drink et libitum, c: Treatment group 1, namely male white rats of the Wistar strain that were induced by gentamicin and received a standard feed diet, drink et libitum and were given tender coconut water at a dose of 8 mL/200g BW for 2 weeks, d: Treatment group 2, namely male white rats of the Wistar strain that were induced by gentamicin and received a standard feed diet, drink et libitum and were given tender coconut water at a dose of 8 mL/200g BW mixed with probiotics at a dose of 18 mg for 2 weeks, e: Treatment group 3, namely male white rats of the Wistar strain that were induced by gentamicin and received a standard feed diet, drink et libitum, tender coconut water at a dose of 8 mL/200g BW mixed with probiotics at dose of 18 mg and vitamin E at a dose of 1.8 IU/200 g BB for 2 weeks.

This was also observed in the group of mice induced by gentamicin and given tender coconut water at a dose of 8 mL/200 g BW only (K3). There were significant differences in SOD and GPx levels between treatment groups 1 and 2, treatment group 1 with treatment group 3, and treatment group 2 with treatment group 3, meaning as follows: There was a significant difference in the average levels of SOD and GPx between group 1, group 2, and 3. There was a significant difference in the average levels of SOD and GPx between treatment 2 (Wistar rats induced by gentamicin) receiving tender coconut water at a dose of 8 mL/200 g BW mixed with probiotics at a dose of 18 mg and vitamin (K4) with treatment group 3, receiving tender coconut water with probiotics at a dose of 8 mL/200 g BW (18 mg) and vitamin E at a dose of 1.8 IU/200 g BW for 2 weeks (K5). The combination of probiotics, tender coconut water, and Vitamin E exhibited a nephroprotective effect in a gentamicin-induced Acute Kidney Injury. The combination exerted an antioxidant effect via an increase in endogenous (SOD and GPx) antioxidant activity. The average levels of SOD and GPx in K5 were the highest, followed by K4 and K3. The lowest levels of SOD and GPx were in group K2 (the group that was only induced by gentamicin). Gentamicin can induce kidney damage and accumulate in the proximal tubule after being filtered in the glomerulus.<sup>2</sup> Accumulation of gentamicin in cells can cause an increase in reactive oxygen species (ROS), which can trigger an increase in free radicals, causing cell damage and kidney necrosis.<sup>3</sup> ROS compounds that act as oxidants are hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), superoxide ion (O<sub>2</sub><sup>•-</sup>), peroxy radical (•OOH), hydroxyl radical (•OH), and singlet oxygen (<sup>1</sup>O<sub>2</sub>).<sup>19</sup> Gentamicin increases the formation of superoxide ions (O<sub>2</sub><sup>•-</sup>), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), and hydroxyl radicals (•OH) and releases iron (Fe) from the mitochondria of the renal cortex. This condition causes *in vitro* lipid peroxidation and is a strong catalyst for forming free radicals. The finding of Patil *et al.* showed an increase in the production of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) in the renal cortex of rats induced by gentamicin.<sup>1</sup>

Recently, gentamicin has also been shown to increase the formation of superoxide ions (O<sub>2</sub><sup>•-</sup>) and hydroxyl radicals (•OH) in the mitochondria of the renal cortex. Superoxide ions (O<sub>2</sub><sup>•-</sup>) are deleterious when present with H<sub>2</sub>O<sub>2</sub> because they will form hydroxyl radicals (•OH), which is the most reactive and hazardous compound.<sup>4</sup> Increased free radicals can cause decreased endogenous antioxidant activity or antioxidant

enzymes such as superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), glutathione (GSH) and glutathione reductase (GR). Endogenous antioxidant activity such as catalase (CAT), superoxide dismutase (SOD), glutathione (GSH), glutathione reductase (GR), and glutathione peroxidase (GPx) can be decreased due to increased ROS formation by gentamicin.<sup>20</sup> Low levels of endogenous antioxidants can be used as a marker for high free radical levels in the body. The mechanism of gentamicin involves the increase in ROS, triggering oxidative stress and causing a decrease in endogenous antioxidants, as shown in Figure 2.

**Figure 2:** Mechanism of gentamicin against ROS, oxidative stress and antioxidant enzymes that trigger Nephrotoxicity.<sup>21</sup>

The body's response to counteract free radicals is by producing endogenous antioxidants. Still, if the free radicals formed exceed endogenous antioxidants, exogenous sources of antioxidants are needed to maintain a physiological balance. Foods containing natural exogenous antioxidants can reduce morbidity and mortality rates, primarily due to oxidative stress, and suppress the high prevalence of degenerative diseases.<sup>19</sup> This study proves that the combination of

probiotics, tender coconut water, and Vitamin E can increase SOD and GPx levels in rats with gentamicin-induced AKI. This effect was observed in the group of rats induced by gentamicin and given tender coconut water at a dose of 8 mL/200 g BW mixed with probiotics at a dose of 18 mg and vitamin E at 1.8 IU/200 g BW (K5) which gave a higher protection (SOD: 66.39% and GPx: 61.94 u/mg levels) when compared to the group of rats induced only by gentamicin (SOD: 23.77% and GPx: 23.77 u/mg) (K2). The group of rats induced by gentamicin and given tender coconut water at a dose of 8 mL/200 g BW mixed with probiotics (K4) showed higher SOD and GPx levels (SOD: 58.20% and GPx: 51.43 u/mg) when compared to the group of rats induced only by gentamicin (K2). The group of rats induced by gentamicin and given tender coconut water at a dose of 8 mL/200 g BW only (K3) showed higher levels of SOD and GPx (SOD: 40.16% and GPx: 39.73 u/mg) when compared to the group of rats induced by gentamicin alone (K2). The highest levels of both SOD and GPx were seen in group (K5), followed by (K4) and (K3). The combination of tender coconut water, probiotics, and vitamin E has a nephroprotective effect on rats in the Acute Kidney Injury model due to gentamicin induction.

A study by Kunle *et al.*<sup>8</sup> stated that tender coconut water protects against kidney and liver damage due to exposure to monosodium glutamate. Zulaikhah *et al.*<sup>17</sup> stated that tender coconut water can increase the levels of antioxidants SOD and GPx in gold miners exposed to mercury. Tender coconut water can increase SOD levels in mice exposed to cigarette smoke.<sup>22</sup> Tender coconut water made into powder and enriched with vitamin E can increase SOD and GPx levels in type 2 DM mice.<sup>23</sup> The active compounds found in tender coconut water and related to antioxidant status are methionine, L-arginine, selenium, vitamin C, and the minerals Zn, Mn, and Cu.<sup>19,24</sup> Methionine is an essential amino acid that contains sulfur. This compound is vital in protein synthesis. Methionine is a sulfur donor for cysteine. Cysteine is a compound the body needs to produce glutathione (GSH). GSH is the most critical antioxidant in the body.<sup>19</sup> The study's results by Azad *et al.* indicate that methionine can prevent nephrotoxicity and eliminate oxidative stress due to gentamicin.<sup>25</sup> Also, in a study Bashan *et al.* stated that administration of L-arginine can protect against kidney damage caused by gentamicin through protection against NO Synthase.<sup>26</sup> L-arginine is a non-essential amino acid in nitric oxide synthase (NO Synthase), a substrate producing citrulline and nitric oxide (NO). NO can inhibit xanthine oxidase (XO), increase SOD levels, total thiol levels (T-SH), vitamin C, and total antioxidants (TAC), and inhibit free radical chain reactions through lipid peroxidation. One of the processes of superoxide formation in the body is the oxidation of xanthine or hypoxanthine catalyzed by xanthine oxidase (XO), producing uric acid and superoxide. Hence, inhibiting the activity of XO by L-arginine leads to superoxide production decrease because the need for SOD to break down superoxide into H<sub>2</sub>O<sub>2</sub> also decreases.<sup>19</sup> In a healthy organism, the endogenous antioxidant system consisting of SOD, CAT, GPx, and GR is responsible for homeostatic metabolism. It controls excess free radicals, but in the nephrotoxic condition caused by gentamicin induction, this system is eroded so that the conversion of H<sub>2</sub>O<sub>2</sub> to hydroxyl radicals (OH•) increases. L-arginine indirectly triggers the activity of the antioxidant enzyme SOD by controlling the formation of H<sub>2</sub>O<sub>2</sub>, which inhibits the accumulation of H<sub>2</sub>O<sub>2</sub>.

Tender coconut water contains selenium at 0.001 mg/100 g. Selenium plays an essential role in protein synthesis and the activity of the enzyme glutathione peroxidase (GPx). GPx activity is greatly influenced by the presence of selenium. Selenium deficiency in the body can reduce GPx activity by up to 90%.<sup>19</sup> The body's selenium needs under normal conditions are met by food, but when the body experiences kidney damage, a greater supply of selenium is needed from exogenous sources. Selenium can protect against kidney damage induced by gentamicin.<sup>4</sup> The effectiveness of SOD enzymes can be increased by the presence of mineral nutrients such as copper (Cu) and zinc (Zn) and manganese (Mn).<sup>19</sup> Increased free radicals due to gentamicin induction can cause decreased SOD activity. The high nutrient content of tender coconut water can be utilized as an essential source of minerals, one of which is as an antioxidant cofactor of SOD. Zulaikhah *et al.* reported that deficiencies of Cu, Zn, and Mn minerals can reduce the activity of Cu-Zn SOD and Mn-SOD. Mn-SOD is a

catalyst for the dismutase reaction of superoxide anion to hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and oxygen (O<sub>2</sub>) in mitochondria. In contrast, Cu-Zn SOD functions as a catalyst for the dismutase reaction of superoxide anion to hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and oxygen (O<sub>2</sub>) in the cytosol.<sup>19</sup> The vitamin C content in tender coconut water makes it effective for increasing antioxidant levels because vitamin C can reduce superoxide radicals, hydrogen peroxide, and reactive oxygen species. The action of vitamin C as an antioxidant indirectly regenerates membrane antioxidant bonds, such as vitamin E, by lifting peroxy radicals and singlet oxygen. Vitamin C works synergistically with vitamin E. Vitamin E, which is oxidized by free radicals into tocopheroxyl radicals, can react with vitamin C after receiving hydrogen ions from vitamin C; it metabolizes to tocopherol (vitamin E) and monodehydroascorbate. Monodehydroascorbate can spontaneously undergo dismutase into ascorbate (vitamin C) and dehydroascorbate.<sup>27</sup> Nugraha *et al.*<sup>13</sup> stated that Vitamin E protects against kidney damage in white mice given ethinyl estradiol. Vitamin E plays a role in preventing ROS and oxidative stress.<sup>28</sup> Probiotics have antioxidant effects by preventing endothelial cell death caused by oxidants.<sup>29</sup> Probiotics can modulate the immune system, resistance to colonization, repair the intestinal barrier, and produce metabolites that can work locally, such as antimicrobials, enzymes, and organic acids. Probiotics have a role in increasing the production of short-chain fatty acids (SCFAs) and inhibiting the production of GDUT (*Gut-Derived Uremic Toxins*) and endotoxins so that probiotics can play a role in maintaining the integrity of the intestinal barrier, reducing the incidence of inflammation, oxidative stress and modulating immunity, thus playing a role in kidney protection.<sup>30</sup> Probiotic strain *Lactobacillus* sp can also repair the small intestine mucosa and increase energy efficiency.<sup>31</sup> Research by Cecilia *et al.* shows that the administration of probiotics can help to sustain healthy intestinal homeostasis, decrease obesity, and promote wellness.<sup>32</sup>

## Conclusion

In this study, rats induced with gentamicin caused Acute Kidney Injury. The combination of probiotics, tender coconut water and Vitamin E has a nephroprotective effect in Acute Kidney Injury model mice due to gentamicin. The combination increased endogenous antioxidants, SOD and GPx levels compared to negative controls. Further studies are needed to evaluate its effectiveness in humans and to explore the underlying biochemical mechanisms in more detail.

## Conflict of Interest

Authors declare no conflict of interest.

## Authors' Declaration

The authors hereby declare that the work presented in this article is original and that any liability for claims relating to the content of this article will be borne by them.

## Acknowledgements

The Directorate of Research, Technology, and Community Engagement (DRTPM) of the Ministry of Education, Culture, Research, and Technology for funding the PTM (Master Thesis Research) grant in the year 2024 under agreement/contract number 108/E5/PG.02.00.PL/2024; 33/B.1/SA-LPPM/VI/2024. The Research and Community Engagement Institute (LPPM) of the Sultan Agung Islamic University (UNISSULA) in Semarang, Central Java, Indonesia. The Integrated Research and Testing Laboratory (LPPT) of Gadjah Mada University (UGM) in Yogyakarta.

## References

- Patil AA, Doijad R, Koparde A. Renoprotective effect of lycopene on renal functional and histopathological changes in gentamycin induced nephrotoxicity in rats. *Res J Pharm Technol.* 2020; 13(7): 3237–3240.
- Zularsil FWR, Loho LL, Lintong MP. Histopathological picture of wistar kidneys given binahong extract after gentamicin administration. *Jurnal e-Biomedik (eBm);* 2016;4(2):1–6.

3. Sujono TA, Rizki FA. Nephroprotective Effect of Ethanolic Extract of Garlic (*Allium sativum* L.) in Gentamicin-Induced Rats. *Pharmakon Jurnal Farmasi Indonesia*. 2020; 1–9.
4. Gamaan M, Zaky H, Ahmed H. Gentamicin-induced nephrotoxicity: A mechanistic approach. *Azhar Int J Pharm Med Sci*. 2023;3(2):11–19.
5. Ardiansyah S, Galuh RH. The Effect of Oral Gentamicin Administration on Uric Acid Levels. *The Journal of Muhammadiyah Medical Laboratory Technologist*. 2018; 2(1): 12–17.
6. Siahaan SG, Lintong MP, Loho LL. Histopathological Features of Wistar Rat (*Rattus norvegicus*) Kidneys Induced with Gentamicin and Treated with Purple Sweet Potato (*Ipomoea batatas* L. Poir)" *Jurnal e-Biomedik*; 2016;4(1): 1-6
7. Zulaikhah ST. The Effect of Tender Coconut Water Administration on Urea Levels in Wistar Rats Exposed to Lead (Pb). *J. Research. Health. Forikes Voice*. 2020; 11(2): 198–201.
8. Kunle-Alabi OT, Akindele OO, Charles KJ, Raji Y. Coconut water prevents renal and hepatic changes in offspring of monosodium glutamate treated wistar rat dams. *Niger. J. Physiol. Sci*. 2019; 34(1): 11–16.
9. Ekezie J, Ndubuka GIN, Ezeiofor TIN. Immature Coconut Water: A Renal Protective Agent in Wistar Rats. *IJAAP*. 2016; 2(2): 26–31.
10. Nwangwa EK. The Reno-Protective Effects of Coconut Water on the Kidneys of Diabetic Wistar Rats. *Int. J. Health Sci*. 2012; 2(1): 1–4.
11. Christanto J, Winata J. Utilizing Coconut Water as a Probiotic Drink Fermented with Lactic Acid Bacteria, *Lactobacillus Bulgaricus*. 2021. Doi 10.17605/OSF.IO/P4E8W
12. Mert H, Mert N, Yoruk M, Yoruk IH, Yildirim BA, Yilmaz HC. Effect of coenzyme Q10 and vitamin E on gentamicin-induced nephrotoxicity in rats. *GSC Biol. Pharm. Sci*. 2022;21(2): 033–040.
13. Nugraha P, Samsuri S, Berata IK. The Effect of Vitamin E and Ethinyl Estradiol Administration on the Histopathological Image of the Kidneys of White Rats (*Rattus Norvegicus*). *Indonesia Medicus Veterinus*. 2018; 7(3):284.
14. Venkatesha U, Veeru P. Gentamicin induced acute renal damage and its evaluation using urinary biomarkers in rats. *Toxicol. Rep*. 2019; 6: 91-99.
15. Nova FS, Chasani S, Hussanna A, Zulaikhah ST. Tender coconut water Inhibits the process of lipid peroxidation, reduce glucose levels, and increase plasma insulin in pregnant diabetic rats. *Pharmacogn J*. 2020; 12(1): 162–167.
16. Zulaikhah ST, Ratnawati R, Hussaana A, Muhandri T. Comparison of Powdered Active Compounds Made from Tender Coconut Water Fortified with Vitamin E, Processed by Spray Drying and Freeze Drying. *Pharmacogn J*. 2022; 14(6): 682–686
17. Zulaikhah ST, Wahyuwibowo J, Suharto MN, Enggartiasto BH, Ortanto MIR, Pratama AA. Effect of tender coconut water (TCW) on TNF- $\alpha$ , IL-1 and IL-6 in streptozotocin (STZ) and nicotinamid (NA) induced diabetic rats. *Pharmacogn J*. 2021; 13(2): 500–505.
18. Dahlan MS. *The Gateway to Understanding Statistics, Methodology and Epidemiology: Multiaxial Sopiudin Dahlan*. Jakarta: Sagung Seto; 2014.
19. Zulaikhah ST, Anies, Santosa AS. Effect of Tender Coconut Water on Antioxidant Enzymatic Superoxida Dismutase (SOD), Catalase (CAT), Glutathione Peroxidase (GPx) and Lipid Peroxidation in Mercury Exposure Workers. *Int. J. Sci. Res*. 2015; 4(12): 517–524.
20. Gamaan M, Zaky H, Ahmed H. Gentamicin-induced nephrotoxicity: A mechanistic approach. *AJPMMS*. 2023; 3(2): 11–19.
21. Abdelrahman RS, Abdelmageed ME. Renoprotective effect of celecoxib against gentamicin induced nephrotoxicity through suppressing NF $\kappa$ B and caspase-3 signaling pathways in rats. *Chem. Biol. Interact*. 2020;315: 108863.
22. Zulaikhah ST, Aini HFN, Rini AS, Abiyyu BH, Dewi EAT, Pratama AA. Tender Coconut Water (*Cocos nucifera* L.) Can Increase Antioxidant Enzymes and Decrease MDA Levels: Experimental Study on Cigarette Smoke-Exposed Rats. *Pharmacogn J*. 2022;14(5): 469–476.
23. Ratnawati R, Zulaikhah ST, Hussaana A, Pratama AA. Effect of Tender Coconut Water Powder Enriched with Vitamin E in Preventing Oxidative Stress of Diabetes Male Wistar Rats. *Pharmacogn J*. 2024; 16(5): 1062–1068.
24. Zulaikhah ST, Sampurna, Wibowo JW, Aini HFN, Pratama AA. Tender Coconut Water Can Inhibit Inflammation Caused by Cigarette Smoke. *J. Hunan Univ. Nat. Sci*. 2021; 48(12): 29-35.
25. Azad MAK, Sivanesan S, Wang J, Chen K, Nation RL, Thompson PE, et al. Methionine ameliorates polymyxin-induced nephrotoxicity by attenuating cellular oxidative stress. *Antimicrob Agents Chemother*. 2018;62(1):1–9.
26. Bashan L, Bashan P, Seçilmis MA, Singirik E. Protective effect of L-arginine on gentamicin-induced nephrotoxicity in rats. *Indian J. Pharmacol*. 2014; 46(6): 608–612.
27. Zulaikhah ST, Pertiwi D, Bagus SA, Nuri S, Alfiza NS. Effect of Tender Coconut Water on Blood Lipid Levels in Hight Fat Diet Fed Male Rats. *J Krishna Inst Medical Sci Univ*. 2017; 6(2): 63–68.
28. Ashrafi S, Heidari R, Ashrafi MR, Chamanara M, Dadpay M, Ebrahimi M. The Protective Effects of Alpha-Tocopherol Against Gentamicin-Induced Nephrotoxicity: The Potential Role of the Nrf2/NQO1 Pathway. *J. Appl. Biotechnol. Rep*. 2024; 11(2): 1334–1343.
29. Patra A, Mandal S, Samanta A, Mondal KC, Nandi DK. Therapeutic potential of probiotic *Lactobacillus plantarum* AD3 on acetaminophen induced uremia in experimental rats. *Clin. Nutr. Exp*. 2018; 19: 12-22.
30. Hsiao WH, Ming J. Exploring the Preventive and Therapeutic Mechanisms of Probiotics in Chronic Kidney Disease through the Gut–Kidney Axis. *J. Agric. Food Chem*. 2024; 72(15): 8347–8364.
31. Zulaikhah ST, Punasari PW. The Effect of Probiotic and Zinc Combination on the Body Weight of Malnourished Rats. *J. Research. Health. Forikes Voice*. 2021; 12(2): 166– 169.
32. Okediya CK, Oyewale JO, Okediya TT, Ajayi AS, Olasehinde GI. Effect of *Lactobacillus acidophilus* and *Lactobacillus plantarum* on weight reduction in obese rats. *Trop J Nat Prod Res*. 2021;5(4):759–762.