



Xanthenes from the Roots of *Garcinia rigida* Miq. and Their Cytotoxic Activity against HeLa Cervical Cancer Lines

Audrey H. Zulfa¹, Salwa Assyifa¹, Ahmad R. Setiawan¹, Mulyadi Tanjung², Tjitjik S. Tjahjandrie², Ratih D. Saputri^{1*}

¹Department of Chemistry, Faculty Mathematics and Natural Sciences, Universitas Negeri Surabaya, Surabaya 60231, Indonesia

²Department of Chemistry, Faculty of Science and Technology, Universitas Airlangga, Surabaya, Indonesia.

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ABSTRACT

Garcinia rigida Miq. (Clusiaceae), a plant endemic to Borneo is a traditional medicine in Kalimantan, Indonesia. Xanthenes are a promising metabolomic substance of *Garcinia* for cancer therapy. This research aimed to evaluate the cytotoxicity against HeLa cells of xanthenes from the roots of *Garcinia rigida* Miq. Two xanthenes, mangostanin (**1**) and 8-isoprenyl-1,6,7-trihydroxy-6',6'-dimethyl-pyrano(2',3':3,2)xanthone (**2**), were isolated from the roots of *G. rigida*. Xanthenes **1-2** molecular structures were established and focused using ¹H, ¹³C, HMQC, and HMBC spectra. Xanthenes **1-2** were assayed regarding their cytotoxicity against HeLa cells using the MTT method. Compound **2** exhibited moderate cytotoxicity with an IC₅₀ of 21.89 μM.

Keywords: *Garcinia rigida*, xanthenes, cytotoxic, HeLa cells.

Introduction

Garcinia rigida Miq. (Clusiaceae) is one endemic plant in the Borneo Island (Indonesia). The genus *Garcinia* produces xanthenes as the main metabolomics that exhibit antioxidant, anti-inflammatory, antimalaria, and cytotoxic activities.¹⁻⁸ Xanthenes show functional groups such as hydroxy, methoxy, and terpenyl chains (isoprenyl, geranyl), which are important in molecular networking to inhibit or kill cancer cells.⁶⁻⁸ Previous studies have shown that xanthenes from *G. cowa* and *G. bracteata* show significant activity against HeLa cells.⁹⁻¹⁰ Two known xanthenes, mangostanin (**1**) and 8-isoprenyl-1,6,7-trihydroxy-6',6'-dimethyl-pyrano(2',3':3,2)xanthone (**2**) were found in the roots of *G. rigida*. The effect of hydroxy groups, isoprenyl chains, and pyrano rings in the structure of xanthenes **1-2** against cervical cancer HeLa cells were also discussed.

Materials and Methods

General experimental procedure

The stationary phases of column chromatography (CC) and planar radial chromatography used silica gel G₆₀ and PF₂₅₄. The TLC plates control the separation process in the CC and planar radial chromatography. The structures of xanthenes **1-2** using a UV spectrophotometer (Thermo Scientific™), an NMR spectrometer (JEOL ECA-400), and FTIR spectrophotometer performed the IR spectrum (FTIR with ATR, Perkin Elmer).

Plant material

The plant material (root) of *G. rigida* originates from Gandring Village (GPS coordinates: 0°93'49" S, 114°89'85" E), Teweh Baru District, South Barito, Central Kalimantan, Indonesia, in March 2023 with the specimen number TB-BJGR-IR5.

Extraction and isolation

For two days, the dried powder of *G. rigida* roots (2.2 kg) was macerated with 90% methanol. The solvents were eliminated using a rotavapor, producing a thick methanol extract weighing 90 grams. The MeOH extract was partitioned with ethyl acetate to produce an ethyl acetate fraction (17.9 g).¹¹⁻¹³

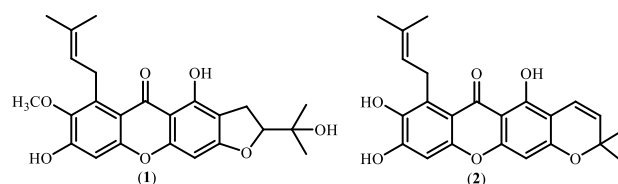


Figure 1: Xanthenes of *G. rigida*

*Corresponding author. E mail: ratihsaputri@unesa.ac.id

Tel: +6231-8298761

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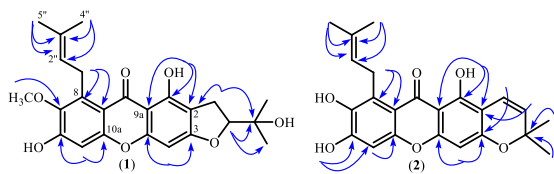


Figure 2: HMBC spectrum of xanthenes 1-2

Silica gel CC was carried out to separate the EtOAc fraction with a gradient of *n*-hexane (C₆H₁₄): EtOAc (18:2 to 1:1 v/v) so that four fractions (A-D) were obtained. By radial chromatography, fraction B (3.3 g) was separated with C₆H₁₄: EtOAc (18:2 to 1:1 v/v) to obtain four subfractions (B₁-B₄). Re-separation of subfraction B₃ (194 mg) by radial chromatography with the eluents diisopropyl ether and diisopropyl ether: ethyl acetate (9:1 v/v) to obtain subfractions B₃₁-B₃₄. Fraction B₃₄ (11.2 mg) was purified using the same method with eluents of *n*-hexane: chloroform (17:3 v/v), chloroform, and chloroform: EtOAc (9:1 v/v) to obtain mangostanin (**1**, 4.9 mg). Subfraction B₁ (25 mg) was separated using C₆H₁₄: chloroform (9:1 v/v) and chloroform to give subfractions B₁₁-B₁₅. Subfraction B₁₄ (14.4 mg) was purified using radial chromatography with C₆H₁₄: acetone (19:1 to 9:1 v/v) eluent,

which gave 8-isoprenyl-1,6,7-trihydroxy-6',6'-dimethyl-pyrano(2',3':3,2)xanthone (**2**, 7.2 mg).

Cytotoxic activity

Xanthenes **1-2** activity assay against cervical cancer (HeLa cells) was performed using the MTT method, and doxorubicin was used as a positive control. The HeLa cells in DMSO without active compounds **1-2** are negative controls in the cytotoxic assay. Compounds **1-2** in various concentrations (1 to 300 μM) and then added to HeLa cell culture. The cells in 96 wells in RPMI-1640 media containing 1.5 g/L Na₂CO₃ were supplemented with 1% L-glutamate, 1% fetal bovine serum, and 1% antibiotic and antifungal formula. Incubation of HeLa cells was carried out in a 5% CO₂ incubator at 37°C for 24 hours. Absorbance was measured using a microplate reader (Diatek DR-200Bc, ELISA, China) at λ_{max} 540 nm.¹⁴⁻¹⁶

Result and Discussion

Two xanthenes (Figure 1), mangostanin (**1**) and 8-isoprenyl-1,6,7-trihydroxy-6',6'-dimethyl-pyrano(2',3':3,2)xanthone (**2**), were found from *G. rigida* roots. The NMR data of xanthenes **1-2** have chemical shift (δ) values identic to those in the literature.²⁰

Table 1: NMR spectra of compounds **1-2** in CDCl₃

No. C	1		2	
	δ _H (mult, J in Hz)	δ _C	δ _H (mult, J in Hz)	δ _C
1	-	158.8	-	158.0
2	-	108.8	-	104.0
3	-	167.9	-	160.0
4	6.25 (s)	88.6	6.24 (s)	94.2
5	6.83 (s)	102.7	6.83 (s)	101.5
6	-	156.2	-	150.9
7	-	144.6	-	139.8
8	-	138.0	-	127.4
9	-	182.9	-	182.8
4a	-	157.5	-	156.4
8a	-	111.8	-	111.6
9a	-	102.7	-	104.4
10a	-	157.9	-	153.7
1'	-	-	-	-
2'	4.81 (dd, 7.3; 9.2)	92.8	-	78.0
3'	3.14 (t, 8.5)	27.0	6.72 (d, 9.8)	127.8
4'	-	71.4	5.56 (d, 9.8)	115.8
5'	1.28 (s)	25.9	1.47 (s)	28.4
6'	1.24 (s)	25.9	1.47 (s)	28.4
1''	4.11 (d, 7.4)	26.8	4.34 (d, 7.3)	26.1
2''	5.26 (t, 6.7)	124.7	5.31 (t, 7.4)	121.4
3''	-	131.4	-	136.0
4''	1.81 (s)	18.2	1.79 (s)	18.2
5''	1.64 (s)	25.5	1.89 (s)	25.9
1-OH	13.67 (s)	-	13.69 (s)	-
5-OH	-	-	5.43 (s)	-
7-OCH ₃	3.79 (s)	61.2	-	-

Table 2: Cytotoxic activity against HeLa cells of xanthenes 1-2.

Compound	μM
1	256.49 \pm 1.05
2	21.89 \pm 0.68
Doxorubicin	1.25 \pm 0.07

The UV (MeOH) spectrum of mangostanin (**1**), displaying λ_{max} (log ϵ): 220 (4.13); 246 (4.26); 278 (4.06); 318 (3.83), and IR (KBr) spectrum shows ν_{max} (cm^{-1}): 3311, 1657, 1573, and 1610.¹⁷

The ^1H NMR of mangostanin displayed 11 total protons of aromatic, chelated hydroxy, methoxy, dihydrofuran rings, and isoprenyl chains (Table 1). At ring A, a singlet aromatic proton exhibits at δ_{H} 6.25 (H-4) and the same signal at δ_{H} 6.83 (H-5) (ring B). Two protons of hydroxy and methoxy groups were detected at δ_{H} 13.67 (1-OH) and δ_{H} 3.79 (7-OCH₃). The proton of the 2'-(1-hydroxy-1-methylethyl)-dihydrofuran chain consists of a methylene proton [δ_{H} 3.14 (H-3')], an oxy-methine [δ_{H} 4.81 (H-2')], and two methyls [δ_{H} 1.24 (H-6'); δ_{H} 1.28 (H-5')]. The proton of the isoprenyl chain consists of a methylene [δ_{H} 4.11 (H-1'')], a vinyl [δ_{H} 5.26 (H-2'')], and two methyls [δ_{H} 1.64 (3H, s, H-5''), δ_{H} 1.81 (3H, s, H-4'')].¹⁸⁻¹⁹ The placement of two aromatic units, hydroxy, methoxy, isoprenyl chain, and 2'-(1-hydroxy-1-methylethyl)-dihydrofuran ring, was detected by HMBC spectrum. The ^{13}C -NMR (Table 1) spectrum of mangostanin (**1**) showed 23 perfectly separated carbon atoms supported by the 2D-NMR spectrum. A hydroxy at 1-OH (δ_{H} 13.67) showed a correlation to an oxyaryl at C-1 (δ_{C} 158.8), and two quaternary carbon [C-2 (δ_{C} 108.8), C-9a (δ_{C} 102.7)]. A methylene proton at H-3' (δ_{H} 3.14) shows cross-peak to C-2 and an oxy-carbon [C-4' (δ_{C} 71.4)]. An aromatic at H-4 (δ_{H} 6.25) linked to C-2, C-9a, and two oxyaryls [C-3 (δ_{C} 167.9), C-4a (δ_{C} 157.5)], indicating on ring A the 2'-(1-hydroxy-1-methylethyl)-dihydrofuran fused at C-2 and C-3. An aromatic at H-5 (δ_{H} 6.83) correlate with three oxyaryls [C-6 (δ_{C} 156.2), C-7 (δ_{C} 144.6), C-10a (δ_{C} 157.9)], C-8a (δ_{C} 111.8, quaternary carbon), 7-OCH₃ (δ_{H} 3.79) linked to C-7, and a methylene proton of the isoprenyl chain at δ_{H} 4.11 (H-1'') correlated to C-7, C-8a, two quaternary carbons [C-8 (δ_{C} 138.0), C-3' (δ_{C} 131.4)], and a methine carbon at C-2' (δ_{C} 124.7) highlighting the isoprenyl chain at C-8 and the methoxy group at C-7. The HMBC correlation shows that ring B is 3-isoprenyl-1-hydroxy-2-methoxybenzene (Figure 2). The xanthone structure of **1** was identified as mangostanin.²⁰

8-Isoprenyl-1,6,7-trihydroxy-6',6'-dimethyl-pyrano(2',3':3,2)xanthone (**2**) has a yellow solid, and the UV spectrum (λ_{max} : 222, 268, 290, and 334 nm) similar to mangostanin. The NMR data of **2** also display the same chemical shift with mangostanin, especially in the two aromatic units [δ_{H} 6.24 (H-4), δ_{C} 94.2 (C-4), δ_{H} 6.83 (H-5), δ_{C} 101.5 (C-5)], an isoprenyl at C-8 [δ_{H} 4.34 (H-1''), δ_{C} 26.1 (C-1''), δ_{H} 5.31 (H-2''), δ_{C} 121.4 (C-2''), δ_{C} 136.0 (C-3''), δ_{H} 1.79 (H-4''), δ_{C} 18.2 (C-4''), δ_{H} 1.89 (H-5''), δ_{C} 25.9 (C-5'')], and the chelated hydroxy at C-1 [δ_{H} 13.69 (1-OH)]. The main difference in compound **2** is that a 2,2-dimethylpyrano ring is linked at C-2 and C-3 [δ_{C} 78.0 (C-2), δ_{H} 6.72 (1H, $J = 9.8$ Hz, H-3'), δ_{C} 127.8 (C-3'), δ_{H} 5.56 (1H, $J = 9.8$ Hz, H-4'), δ_{C} 115.8 (C-4'), δ_{H} 1.47 (6H, s, H-5'/H-6'), δ_{C} 28.4 (C-5'/C-6')], and demethylation at C-7. Twenty-two carbon atom signals in compound **2** were seen in the ^{13}C -NMR spectrum and were perfectly separated (Table 1). The HMBC spectrum of **2** also shows a similar correlation with **1**, especially in two aromatic units, an isoprenyl at C-8 and the hydroxy group at C-1. The hydroxy proton at 1-OH correlated to C-1 (δ_{C} 158.0, oxyaryl), C-2, and C-9a (δ_{C} 104.0, δ_{C} 104.4, quaternary carbon). A vinylic proton at H-4' (δ_{H} 5.56) correlates with C-2', and C-3 emphasizes the pyrano ring linked at C-2 and C-3. Based on the NMR spectrum, the structure **2** is as shown in Figure 1.²⁰

Xanthenes **1-2** showed IC₅₀ values of 256.49 and 21.89 μM , respectively (Table 2) against HeLa cells. Compound **2** described moderate activity, and mangostanin was inactive.²¹⁻²² From the structure, compound **2** has a 2,3-dimethylpyrano ring compared to compound **1**, which shows a dihydrofuran ring connected at C-2 and C-3, and demethylation at C-7 increases activity. The planarity of the 2,3-dimethylpyrano ring can penetrate and damage HeLa cells so that its activity increases.

Conclusion

Two known xanthenes, mangostanin (**1**) and 8-isoprenyl-1,6,7-trihydroxy-6',6'-dimethyl-pyrano(2',3':3,2)xanthone (**2**), were found in the roots of *G. rigida* Miq. Compound **1** was categorized as inactive against HeLa cells, and compound **2** had moderate activity.

Conflict of Interest

The author and all members of the research team declare that there is no conflict of interest.

Author's Declaration

The author and research team state that the research is original, and the content of this article is the author's responsibility.

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References

- Othman NA, Liew SY, Khaw KY, Ablat A, Karsani SA, Leong KH, Blanchard P, Derbré S, Awang K. Secondary metabolites from the barks of *Garcinia griffithii* T. Anderson (Clusiaceae) and their chemophenetic significance. *Biochem. Syst. Ecol.* 2024; 114: 104818.
- Mustafa NH, Jalil J, Leong KE, Jamal JA, Husain K. Phytochemical profile and diverse *Garcinia celebica* L. *Heliyon* . 2024; 10. e30629.
- Abate M, Pagano C, Masullo M, Citro M, Pisanti S, Piacente S, Bifulco M. Mangostanin, a xanthone derived from *Garcinia mangostana* fruit, exerts protective and reparative effects on oxidative damage in human keratinocytes, *Pharmaceuticals*, 2022; 15(1): 84.
- Fadlan RA, Fatmawati S, Purnomo AS, Ersam T. Antiplasmodial and anticancer activities of xanthenes isolated from *Garcinia bancana* Miq. *Nat. Prod. Res.* 2023; 1-6.
- Sukandar ER, Kaennakam S, Raab P, Nöst X, Rassamee K, Bauer R, Siripong P, Ersam T, Tip-Pyang S, Chavasiri W. Cytotoxic and anti-inflammatory activities of dihydroisocoumarin and xanthone derivatives from *Garcinia picrorhiza*. *Molecules*. 2021; 26(21), 1-11.
- Zhang X, Song Z, Li Y, Wang H, Zhang S, Reid AM, Lall N, Zhang J, Wang C, Lee D, Ohizumi Y, Xu J, Guo Y. Cytotoxic and antiangiogenic xanthenes inhibiting tumor proliferation and metastasis from *Garcinia xipshuanbannaensis*. *J. Nat. Prod.* 2021; 84(5), 1515–1523.
- Ibrahim SRM, Mohamed GA, Elfaky MA, Al Haidari RA, Zayed MF, El-Kholy AAE, Khedr AIM. Garcixanthone A, a new cytotoxic xanthone from the pericarps of *Garcinia mangostana*. *J. Asian Nat. Prod. Res.* 2019; 21(3), 291–297.
- Jia C, Xue J, Gong C, Li X, Li D, Li Z, Hua H. Chiral resolution and anticancer effect of xanthenes from *Garcinia paucinervis*. *Fitoterapia*. 2018; 127, 220–225.
- Zhang B-J, Fu W-W, Wu R, Yang J-L, Yao C-Y, Yan B-X, Tan H-S, Zheng C-W, Song Z-J, Xu H-X. Bioactive scalemic caged xanthenes from the leaves of *Garcinia bracteata*. *Bioorg. Chem.* 2019; 82, 274-283.

10. Kaennakam S, Siripong P, Tip-pyang S. Kaennacowanols A-C, three new xanthenes and their cytotoxicity from the roots of *Garcinia cowa*. *Fitoterapia*. 2015; 102, 171-176.
11. Aminah NS, Kristanti AN, Tanjung M. Antioxidant activity of flavonoid compounds from the leaves of *Macaranga gigantea*. *J. Chem. Pharm. Res.* 2014; 6(6), 688-692.
12. Tanjung M, Saputri RD, Tjahjandarie TS. Antioxidant activity of two isomeric benzoxepin derivatives from the stem bark of *Bauhinia aculeata* L. *J. Chem. Pharm. Res.* 2014; 6(1), 705-708.
13. Tjahjandarie TS, Tanjung M, Saputri RD, Nadar PB, Aldin MF, Marlina E, Permadi A. Flavestins K, a new isoprenylated stilbene from the leaves of *Macaranga recurvata* Gage. *Nat. Prod. Sci.* 2019; 25(3), 244-247.
14. Tanjung M, Nurmalsari I, Wilujeng AK, Saputri RD, Rachmadiarti F, Tjahjandarie TS. Acronyculatin P, a new isoprenylated acetophenone from the stem bark of *Acronychia pedunculata*. *Nat. Prod. Sci.* 2018; 24(4): 284-287.
15. Tjahjandarie TS, Tanjung M, Saputri RD, Rahayu DO, Gunawan ANI, Aldin MF, Aldin MF. Two new 2-arylbenzofurans from *Sesbania grandiflora* L. and their cytotoxicity towards cancer cells. *Nat. Prod. Res.* 2021; 35(24): 5637-5642.
16. Saputri RD, Tjahjandarie TS, Tanjung M. Two novel coumarins bearing acetophenone derivative from the leaves of *Melicope quercifolia*. *Nat. Prod. Res.* 2021; 35(8): 1256-1261.
17. Saputri RD, Tukiran, Suyatno, Wati FA, Dzulkarnain SA, Zakiah M, Tjahjandarie TS, Tanjung M. Xanthine oxidase inhibitory activity of xanthenes from *Calophyllum pseudomole* P.F. Stevens, *Trop. J. Nat. Prod. Res.* 2024; 8(1): 5932-5935.
18. Tanjung M, Tjahjandarie TS, Saputri RD, Kurnia BD, Rachman MF, Syah YM. Calotetrapterins A-C, three new pyranoxanthenes and their cytotoxicity from the stem bark of *Calophyllum tetrapterum*. *Nat. Prod. Res.* 2021; 35(3): 407-412.
19. Mardhiyyah S, Zakiah M, Renata ED, Tjahjandarie TS, Supratman U, Retnowati R, Saputri RD, Tanjung M. Cytotoxic activity of xanthenes from the stem bark of *Cratoxylum sumatranum*, *Trop. J. Nat. Prod. Res.* 2023; 7(9): 3908-3910.
20. Suksamrarn S, Komutiban O, Ratananukul P, Chimnoi N, Lartpornmatulee N, Suksamrarn A. Cytotoxic prenylated xanthenes from the young fruit of *Garcinia mangostana*. *Chem. Pharm. Bull.* 2006; 54(3): 301-305.
21. Tanjung M, Tjahjandarie TS, Aldin MF, Mardhiyyah S, Ahmat N, Saputri RD. Macagigantin A, a new flavonoid from *Macaranga gigantea* (Rchb.f & Zoll). *Mull. Arg. Nat. Prod. Sci.* 2023; 29(4): 287-290.
22. Nurlelasari Rahmayanti I, Salam S, Safari A, Harneti D, Maharani R, Hidayat AT, Tanjung M, Retnowati R, Shiono Y, Supratman, U. A new havanensin-type limonoid from *Chisocheton macrophyllus*. *Appl. Biol. Chem.* 2021; 64: 35.