Tropical Journal of Natural Product Research

Available online at https://www.tjnpr.org



Review Article

Acacetin and Chrysoeriol: A Short Review of the Chemistry, Plant Sources, Bioactivities and Structure-Activity Relationships of these Methylated Flavones

Eric W. C. Chan¹*, Siu K. Wong², Hung T. Chan³

¹Faculty of Applied Sciences, UCSI University, 56000 Cheras, Kuala Lumpur, Malaysia ²Xiamen University Malaysia, Bandar Sunsuria, 43900 Sepang, Selangor, Malaysia ³Secretariat of International Society for Mangrove Ecosystems (ISME), Faculty of Agriculture, University of the Ryukyus, Okinawa 903-0129, Japan

ARTICLE INFO

ABSTRACT

Article history: Received 23 September 2021 Revised 17 November 2021 Accepted 10 December 2021 Published online 03 February 2022

Copyright: © 2022 Chan *et al.* This is an openaccess article distributed under the terms of the <u>Creative Commons</u> Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Flavonoids are plant secondary metabolites that are well-known for their bioactivities. In this article, the chemistry, plant sources, bioactivities and structure-activity relationships of acacetin (ACT) and chrysoeriol (CSE) are reviewed. Of these two flavones, only ACT has been reviewed but not CSE. Sources of information cited were from Google Scholar, PubMed, PubMed Central, Science Direct, Web of Science, J-Stage, PubChem and Directory of Open Access Journals (DOAJ). The criteria used for selection of articles are based on topics rather than period of coverage, although recent references accord higher priority. Flavones, a class of flavonoids, have a C2–C3 double bond and a 4-carbonyl group but lack the C3 hydroxyl group at ring C. ACT and CSE are lesser-known methylated flavones with hydroxyl groups at C5 and C7 of ring A. The methoxy group of ACT is at C4' while that of CSE is at C3'. Found in many plant species, ACT and CSE have generated much research interest because of their diverse pharmacological activities, notably, their anti-cancer properties. The anti-cancer effects and molecular mechanisms of ACT towards lung, liver, gastric, prostate, breast and squamous cancer cells including leukaemia have been reported. Studies have shown that CSE inhibited breast, lung and pancreatic cancer cells including myeloma. Other bioactivities and structure-activity relationships of ACT and CSE are also briefly mentioned. Some areas for further research are suggested.

Keywords: Acacetin, Chrysoeriol, Methylated Flavones, Cytotoxicity, Structure-Activity Relationships.

Introduction

Flavonoids are the largest family of phenolic secondary metabolites, found in almost all herbs, fruits and vegetables.¹⁻³ Their molecular structures consist of two benzene rings A and B that are joined by a heterocyclic pyran ring C forming the benzo-pyrone (C6-C3–C6) moiety.^{4,5} Rings A and C compose of the chroman (C6–C3) nucleus.⁶ Flavonoids can be divided into classes such as flavones, flavonols, flavanones and flavanols.^{4,5,7} Flavones (examples: apigenin and luteolin) have a C2-C3 double bond and a 4-carbonyl group, but lack the C3 hydroxyl group at ring C. Flavonols (e.g., fisetin, quercetin, morin and myricetin) possess all the three functional moieties. Flavanones (e.g., naringenin, hesperitin and taxifolin) lack the C2-C3 double bond while flavanols (e.g., catechin and epicatechin) lack the C2–C3 double bond and the 4-carbonyl group.¹ Flavonoids have been reported to possess broad bioactivities such as anticancer, immunomodulation and antioxidant activities, that can be enhanced, to a certain extent, by methylation.⁸ Methylated flavones containing only one or two methoxy groups are metabolically more stable than polymethoxylated flavones and have more superior chemopreventive properties.9 Investigations on the structure-activity

*Corresponding author. E mail: <u>chanwc@ucsiuniversity.edu.my;</u> <u>erchan@yahoo.com</u> Tel: +603-9101 8880

Citation: Chan EWC, Wong SK, Chan HT. Acacetin and Chrysoeriol: A Short Review of the Chemistry, Plant Sources, Bioactivities and Structure-Activity Relationships of these Methylated Flavones. Trop J Nat Prod Res. 2022; 6(1):1-7 doi.org/10.26538/tjnpr/v6i1.1

Official Journal of Natural Product Research Group, Faculty of Pharmacy, University of Benin, Benin City, Nigeria.

relationship of polymethoxylated flavones such as nobiletin and tangeretin showed a correlation between the number/position of methoxy group and their antiproliferative activity.^{10,11}

In this short review, the chemistry, plant sources, bioactivities and structure-activity relationships of acacetin (ACT) and chrysoeriol (CSE) are reviewed. These methylated flavones are found in many plant species and have been reported to exhibit diverse pharmacological properties notably anti-cancer activities. To date, only ACT has been reviewed, ^{12,13} while CSE has not been reviewed.

Chemistry

Acacetin

ACT (5,7-dihydroxy-4'-methoxyflavone) is a natural methylated flavone.¹² Its molecular formula is $C_{16}H_{12}O_5$ and its molecular weight is 284 g/mol. Being a flavone, ACT has a C2–C3 double bond, a 4-carbonyl group but lacks the C3 hydroxyl group of ring C (Figure 1).

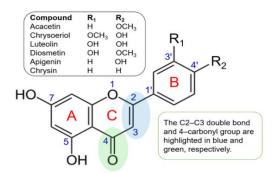


Figure 1: Molecular Structures of Acacetin, Chrysoeriol, Luteolin, Diosmetin, Apigenin and Chrysin

The molecule has two hydroxyl groups at C5 and C7, and one methoxy group at C4'. ACT is also known as apigenin-4'-methyl ether because it is structurally similar to apigenin which lacks the 4'-methoxy group. ACT is also similar to diosmetin, another methoxyflavone with hydroxyl groups at C3', C5 and C7, and a C4'-methoxy group.¹⁴

Chrysoeriol

CSE (4',5,7-trihydroxy-3'-methoxyflavone) is another natural methylated flavone. Its molecular formula is $C_{16}H_{12}O_6$ and its molecular weight is 300 g/mol. Being a flavone, CSE has a C2–C3 double bond, a 4-carbonyl group but lacks the C3 hydroxyl group of ring C (Figure 1). The molecule has three hydroxyl groups at C5, C7 and C4', and one methoxy group at C3'. CSE is also called luteolin-3'-methyl ether because it is structurally similar to luteolin which lacks the 3'-methoxy group. CSE is similar to diosmetin in that they both have a methoxy group and a hydroxyl at the B ring i.e., C3' and C4' for CSE and C4' and C3' for diosmetin.

Plant Sources

Acacetin

ACT is found in the aerial parts of *Chrysanthemum zawadskii* (*Zawadskii* chrysanthemum),¹⁵ *Agastache mexicana* (Mexican giant hyssop),¹⁶ *Potentilla evestita* (cinquefoil),¹⁷ *Ziziphora clinopodioides* (blue mint bush),¹⁸ *Robinia pseudoacacia* (black locust tree),¹⁹ *Artemisia* species (mugworts),²⁰ and *Saussurea involucrata* (snow lotus);²¹ flowers of *Chromolaena odorata* (Siam weed),²² and *Chrysanthemum morifolium* (florist's daisy);²³ seeds of *Carthamus tinctorius* (safflower);²⁴ and *Acacia* honey.²⁵ In the review on the

therapeutic potential of ACT,¹³ plant sources of ACT consist of 80 plant species, commonly reported in the genera of *Artemisia* (five species) and *Chrysanthemum* (four species).

Chrysoeriol

CSE is found in various plant species including the flowers of *Lonicera japonica* (Japanese honeysuckle),²⁵ tea of *Aspalathus linearis* (rooibos),^{27,28} aerial parts of *Medicago sativa* (alfafa),²⁹ leaves of *Eurya ciliata* (no common name),^{30,31} and flowers of *C. morifolium* (florist's daisy).³²

Anti-Cancer Properties

Acacetin

ACT inhibited the growth of A549 lung and MCF-7 breast cancer cells with IC₅₀ values of 9.46 μ M³³ and 26.4 μ M,³⁴ respectively. ACT from the aerial parts of *C. zawadskii* showed strong cytotoxic activity against HCT116 colon and UO-31 renal cancer cells with IC₅₀ values of 2.44 and 2.89 μ g/mL, respectively.¹⁵ Luteolin, the other compound isolated, displayed no activity against the tested cancer cells. Tested against Jurkat T leukaemia and HSC-3 oral squamous carcinoma cells, the IC₅₀ values of ACT were 25.8 μ M³⁵ and 25.0 μ g/mL,³⁶ respectively. The anti-cancer effects and molecular mechanisms of ACT towards different lung (A549), liver (HepG2), gastric (AGS), prostate (LNCaP and DU145), breast (MCF-7), oral squamous (HSC-3), head and neck squamous (UM-SCC-22A), and colon (SW480 and HCT-116) cancer cells including leukaemia (Jurkat T and B-lymphocytes), FaDu pharyngeal carcinoma, and U87 glioblastoma are shown in Table 1.

Table 1: Anti-Cancer Effects and Molecular Mecha	nisms of Acacetin towards Different Cancer Cell Lines
--	---

Cancer cell line & type	Anti-cancer effect and molecular mechanism of ACT	Reference
A549 lung	Induced cell cycle arrest at G1 phase and cell apoptosis involving the expression	33
	of p53 and activity of the Fas/Fas ligand.	
MCF-7 breast	Induced apoptosis via caspase cascade, mitochondria-mediated death signalling	34
	and SAPK/JNK1/2-c-Jun activation.	
Jurkat T leukaemia	Induced apoptosis <i>via</i> up-regulation of Bax, down-regulation of Bcl-2, and possibly by activation of	35
	the Fas-mediated pathway.	24
HSC-3 oral squamous	Induced apoptosis through activation of a MAPK-mediated pathway followed by induction of a mitochondria- and caspase-dependent mechanism.	36
HepG2 liver	Inhibited cell proliferation by arresting cell cycle progression and induced apoptosis involving the	37
	activity of p53 and Fas/Fas ligand.	
AGS gastric	Triggered apoptosis was mainly associated with ROS production, mitochondrial dysfunction, and Fas activation.	38
LNCaP & DU145	Inhibited cell proliferation and cell cycle progression, and induced apoptotic cell death accompanied	39
prostate	by PARP cleavage.	
A549 lung	Inhibited cell proliferation by reducing MMP-2 and u-PA expressions via reduced phosphorylation of	40
-	JNK, and reducing NF-κB and AP-1 binding activities.	
DU145 prostate	Inhibited cell invasion and migration via inactivation of the p38 MAPK signalling pathway.	41
A549 lung	Inhibited cell invasion and migration via inactivation of p38α and involvement of	42
	the MKK and/or MLK signalling pathways.	
DU145 prostate	Exhibited in vitro and in vivo anti-cancer activity via the suppression of NF-KB/Akt signalling.	43
B-lymphocytes	Induced apoptosis by targeting mitochondria, through increased ROS formation, MMP collapse,	44
eukaemia	increased MPT, release of cytochrome c and caspase 3 activation.	
UM-SCC-22A head & neck	Induced apoptosis via cytochrome c release, activation of caspase-3, and possibly involving the	45
squamous	muscarinic M3R pathway.	
FaDu pharyngeal carcinoma	Inhibited cell growth and induced apoptosis via the death receptor-mediated and	46
	the mitochondria-mediated apoptotic pathways.	
SW480 & HCT-116 colon	Induced mitochondrial ROS-mediated cell death by inducing AIF.	47
U87 glioblastoma	Induced Cdk-cyclin mediated G2/M phase arrest and triggered ROS-mediated apoptosis.	48

Abbreviations: AIF = apoptosis-inducing factor, AP-1 = activator protein 1, Bax = Bcl-2 associated X protein, Bcl-2 = B-cell lymphoma 2, JNK = c-jun N-terminal kinase, M3R = M3 receptor, MAPK = mitogen-activated protein kinase, MKK = mitogen-activated protein kinase, MLK = mixed-lineage protein kinase, MMP = matrix metallopeptidase, MPT = mitochondrial permeability transition, NF- κ B = nuclear factor-kappa B, PARP = poly-(ADP-ribose) polymerase, ROS = reactive oxygen species, SAPK = stress-activated protein kinase, and u-PA = urokinase-type plasminogen activator.

Chrysoeriol

CSE inhibited the proliferation of RPMI 8226 and KM3 multiple myeloma cells at IC₅₀ values of 26 and 35 μ mol/L, respectively.⁴⁹ Against leukaemia HL-60 cells, growth inhibitory effects of CSE were dose-dependent with IC₅₀ value of 29 μ M⁵⁰ and 15 μ M against A549 lung cancer cells.⁵¹ Against MRC-5 normal lung cells, cytotoxicity of CSE was significantly weaker with IC₅₀ value at 93 μ M.⁵¹ Earlier, the anti-proliferative activity of CSE, tested against MCF-7 breast, DMS-114 lung, HT-29 colon, SL-MEL5 melanoma, DU-145 prostate cancer cells displayed IC₅₀ values of 7, 17, 20, 23 and 30 μ M, respectively.⁵² In comparison, luteolin (CSE without the methoxy group) exhibited IC₅₀ values of 3, 11, 21, 32 and 32 μ M, respectively.

In recent years, several studies reported on the anti-cancer properties of CSE by testing different cancer cell lines. CSE significantly inhibited cell proliferation and regulated cell cycle of RPMI 8266 and KM3 multiple myeloma cells by suppression of the PI3K-AKT-mTOR pathway.⁴⁹ CSE promoted cell cycle arrest at G2/M and inhibited migration and invasion of MDA-MB-231 cells by down-regulation of matrix metallopeptidase 9 (MMP-9) and cyclooxygenase-2 (COX-2) expression.⁵³ CSE exerted *in vitro* and *in vivo* cytotoxic effects on A549 lung cancer cells *via* the activation of autophagy, sub-G1 cell cycle arrest, cell migration and invasion inhibition, and modulation of the mitogen-activated protein kinase (MAPK)/extracellular signal-regulated kinase (ERK) signalling pathway.⁵¹ 8-CSE triggered apoptosis of SW1990 pancreatic cancer cells *in vitro* by inhibiting B-cell lymphoma 2 (BCL-2), the anti-apoptotic protein.⁵⁴

Related to anti-cancer activities, CSE inhibited DNA adduct formation with benzo[α]pyrene⁵⁵ and inhibited the formation of a carcinogenic estrogen metabolite⁵⁶ in MCF-7 breast cancer cells. In addition, CSE inhibited the efflux transporter breast cancer resistance protein (BCRP/ABCG2) more strongly than ACT with IC₅₀ values of 0.01 and 0.14 μ M, respectively.⁵⁷ The anti-cancer effects and molecular mechanisms of CSE towards different breast (MCF-7 and MDA-MB-231), pancreas (SW1990), lung (A549) cancer cells including myeloma (RPMI 8226 and KM3) are shown in Table 2.

Other Bioactivities

Acacetin

In this short review, the bioactivities of ACT are updated based on recent reviews.^{12,13} ACT possesses antibacterial,⁵⁹ antiviral,⁶⁰ anti-diabetic,⁶¹ anti-neuroinflammatory,^{62,63} anti-arthritic,⁶⁴ anti-aging,⁶⁵ anti-Alzheimer⁶⁶ and antinociceptive⁶⁷ properties (Table 3).

Chrysoeriol

The bioactivities of CSE have been briefly described in recent reviews.^{68,69} CSE displays lipase inhibitory,⁷⁰ antibacterial,⁷¹ antiinflammatory,⁷²⁻⁷⁴ anti-diabetic⁷⁵ and neuroprotective⁷⁶ activities (Table 3).

Structure-Activity Relationships

Acacetin

Results of a structure-activity relationship (SAR) study showed that ACT had no 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging activity (IC₅₀ value > 500 μ M) while luteolin displayed strong activity (IC₅₀ value of 17.8 μ M).⁷⁷ ACT and luteolin are flavones with hydroxyl groups at C5 and C7 of ring A (Figure 1). ACT has a methoxy group at C4' while luteolin has hydroxyl groups at C3' and C4' of ring B. This indicated that the methoxy group of ACT attributed to its effective cytotoxic activity against cancer cells. A SAR study showed that linarin and linarin acetate with a rhamnose substitution at C7 displayed much weaker induction of apoptosis than ACT with a hydroxyl group at C7.³⁹

Chrysoeriol

Unlike luteolin that displayed strong DPPH radical scavenging activity $(IC_{50} \text{ value} = 17.8 \ \mu\text{M})$, CSE showed no such activity $(IC_{50} \text{ value} > 500 \ \mu\text{M})$.⁷⁷ Results of a SAR study for inhibition of matrix metallopeptidase-9 (MMP-9) and cyclooxygenase-2 (COX-2) activity by the flavonoids showed that flavones had better inhibitory activities when compared to flavonols.⁵³ CSE was found to be the most active followed by ACT, diosmetin and luteolin and apigenin (Figure 1). CSE with substitutions such as 5,7-dideoxychrysoeriol (lacking OH groups at C5 and C7) and 2,3-dihydrochrysoeriol (lacking the C2-C3 double bond), showed weaker inhibition of MMP-9 and COX-2 activity than CSE.⁵³ With regard to proliferation of HL-60 leukaemia cells by 5,7-dihydroxyflavones, diosmetin and CSE induced ~80% inhibition.50 The inhibitory effects of chrysin, apigenin, acacetin and luteolin were weaker. It was suggested that the combinations of hydroxyl and methoxy groups at the B ring influenced the inhibitory effects of the compounds on HL-60 cell proliferation. A recent study on CSE and its analogues showed that they inhibited both mesenchymal-epithelial transition factor (c-Met) and vascular endothelial growth factor receptor 2 (VEGFR2) that are involved in tumorigenesis of certain types of cancer.78 To address cancer drug resistance, further SAR analyzes are needed to guide structural optimizations.

Table 2: Anti-Cancer Effects and Molecular Mechanisms of Chrysoeriol towards Different Cancer Cell Lines

Cancer cell line & type	Anti-cancer effect and molecular mechanism of CSE	Reference
RPMI 8226 & KM3	Inhibited cell proliferation by regulation of cell cycle and inhibition of the PI3K-AKT-mTOR pathway.	49
myeloma		
A549 lung	Exerted in vitro and in vivo cytotoxic effects through activation of autophagy, sub-G1 cell cycle arrest, cell	51
	migration and invasion inhibition via inhibition of the MAPK/ERK signalling pathway.	
MDA-MB-231 breast	Promoted cell cycle arrest at G2/M, and inhibited migration and invasion of cells by by down-regulation of	53
	MMP-9 and (COX-2) expression via the NF-κB pathway.	
SW1990 pancreas	Triggered cell apoptosis by inhibiting BCL-2, the anti-apoptotic protein.	54
MCF-7 breast	Inhibited DNA adduct formation with $benzo[\alpha]$ pyrene in cancer cells.	55
MCF-7 breast	Inhibited the formation of carcinogenic estrogen metabolite in cancer cells.	56
MCF-7 breast	Inhibited TNFa-induced CYP19 expression through the inhibition of ERK1/2-mediated EGR-1 expression.	58

Abbreviations: BCL-2 = B-cell lymphoma 2, COX-2 = cyclooxygenase-2, CYP19 = cytochrome P450 19, EGR-1 = early growth response gene 1, ERK1/2 = extracellular signal-regulated kinase 1/2, MAPK = mitogen-activated protein kinase, MMP-9 = matrix metallopeptidase 9, mTOR = mechanistic target of rapamycin, NF- κ B = nuclear factor-kappa B, PI3K = phosphoinositide 3-kinases, and TNF α = tumor necrosis factor alpha.

Table 3: Other Bioactivities and Mechanisms of Acacetin (ACT) and Chrysoeriol (CSE)

Flavone	Bioactivity	Effect and mechanism	Reference
ACT	Antibacterial	ACT inhibited in vitro and in vivo virulence factor of Gram-positive Streptococcus pneumoniae	59
Anti-o Anti- Anti-a Anti-a Anti-a		by targeting spore-forming activity or pneumolysin.	
	Antiviral	ACT is a potent inhibitor of replication of HSV1.	60
	Anti-diabetic	ACT enhanced glucose uptake through insulin-independent GLUT4 translocation in L6	61
		myotubes.	
	Anti- neuroinflammatory	ACT attenuated LPS-induced neuroinflammation in mice by suppressing microglial activation	62
		and reducing neuronal cell death.	
		ACT protected dopaminergic cells against MPTP-induced neuroinflammation in vitro and in	63
		vivo.	
	Anti-arthritic	ACT displayed anti-arthritic effects in FLS cells.	64
	Anti-aging	ACT promoted healthy aging in Caenorhabditis elegans by altering stress response.	65
	Anti-Alzheimer	ACT protected against Aß production (target of Alzheimer's disease treatment) by reducing	66
		APP protein expression and BACE-1 activity, and inhibited APP synthesis that resulted in a	
		decrease in the number of amyloid plaques.	
	Antinociceptive	ACT decreased visceral and inflammatory nociception, and prevented formalin-induced	67
		oedema in pain-related diseases.	
CSE	Anti-diabetic	CSE exhibited anti-diabetic properties by inhibition of the activity of lipase.	70
		CSE ameliorated hyperglycaemia in streptozotocin-induced diabetic rats by regulating	71
		carbohydrate metabolic enzymes.	
	Antibacterial	CSE exhibited antibacterial activity against nine pathogens at 40 μg per disc, and had a MIC	72
		value of 1.25 µg/mL against MRSA.	
	Anti-inflammatory	The inhibitory effects of CSE on AP-1 activation may be associated with its potent NO	73
		blocking and anti-inflammatory activity.	
		CSE ameliorated TPA-induced skin inflammation in mice by inhibiting NF- κ B and STAT3	74
		pathways.	
		CSE ameliorated COX-2 expression in LPS-stimulated murine macrophages through NF- κB ,	75
		AP-1 and MAPK regulation.	
	Neuroprotective	CSE mediated mitochondrial protection in MPP ⁺ -treated SH-SY5Y cells (a typical in vitro PD	76
		model) via PI3K/Akt pathway.	

Abbreviations: AB = B-amyloid, AD = Alzheimer's disease, AP-1 = activator protein 1, $\overline{APP} = amyloid precursor protein$, BACE-1 = amyloid precursor cleaving enzyme, COX-2 = cyclooxygenase-2, FLS = fibroblast-like synoviocyte, GLUT-4 = glucose transporter type 4, HSV1 = herpes simplex virus type-1, LPS = lipopolysaccharide, MAPK = mitogen-activated protein kinase, MIC = minimum inhibitory concentration, $MPP^+ = 1$ -methyl-4-phenylpyridinium iodide, MPTP = 1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine, MRSA = methicillin-resistant*Staphylococcus aureus*, $NF-\kappa B =$ nuclear factor-kappa B, NO = nitric oxide, PD = Parkinson's disease, PI3K = phosphoinositide 3-kinases, STAT3 = signal transducers and activators of transcription 3, and TPA = 12-*O*-tetradecanoylphorbol-13-acetate.

Conclusion

Flavonoids are a very large plant family of phenolic secondary metabolites. These compounds have a molecular structure consisting of two benzene rings A and B joined by a pyran ring C to form a benzo-pyrone (C6–C3–C6) moiety. The majority of flavonoids have the B ring linked in position 2 to the C ring, and they can be further divided into classes such as flavones, flavonols, flavanols and flavanones. Among the flavones, SAR studies have shown that the presence of the C2–C3 double bond and the 4-carbonyl group at ring C, the absence of the C3 hydroxyl group at ring C, and the pattern of hydroxylation at ring B are associated with enhanced cytotoxicity towards cancer cells. ACT and CSE are methylated flavones with hydroxyl groups at C5 and C7 of ring A. ACT has a methoxy group at C3'. Further research on the structural modifications of ACT and CSE

is needed to synthesis novel derivatives with enhanced anti-cancer properties. Clinical research on ACT and CSE is warranted to evaluate their safety and chemopreventive efficacy when used alone or in combination with other chemotherapy agents.

Conflict of Interest

The authors had no conflict of interest.

Authors' Declaration

The authors hereby declare that this short review is original and that any liability for claims relating to the contents will be borne by them.

References

- Panche AN, Diwan AD, Chandra SR. Flavonoids: An overview. J Nutr Sci. 2016; 5:1-15.
- Guven H, Arici A, Simsek O. Flavonoids in our foods: A short review. J Basic Clin Health Sci. 2019; 3:96-106.
- Kopustinskiene DM, Jakstas V, Savickas A, Bernatoniene J. Flavonoids as anticancer agents. Nutrients. 2020; 12(2):457.
- Singh M, Kaur M, Silakari O. Flavones: An important scaffold for medicinal chemistry. Eur J Med Chem. 2014; 84:206-239.
- Raffa D, Maggio B, Raimondi MV, Plescia F, Daidone G. Recent discoveries of anticancer flavonoids. Eur J Med Chem. 2017; 142:213-228.
- Kanadaswami C, Lee LT, Lee PP, Hwang JJ, Ke FC, Huang YT, Lee MT. The antitumor activities of flavonoids. *In Vivo*. 2005; 19(5):895-909.
- Wang TY, Li Q, Bi KS. Bioactive flavonoids in medicinal plants: Structure, activity and biological fate. Asian J Pharm Sci. 2018; 13(1):12-23.
- Wen L, Jiang Y, Yang J, Zhao Y, Tian M, Yang B. Structure, bioactivity, and synthesis of methylated flavonoids. Ann N Y Acad Sci. 2017; 1398(1):120-129.
- 9. Walle T. Methoxylated flavones, a superior cancer chemopreventive flavonoid subclass? Sem Cancer Biol. 2007; 17(5):354-362.
- Kawaii S, Ishikawa Y, Yoshizawa Y. Relationship between the structure of methoxylated and hydroxylated flavones and their antiproliferative activity in HL60 cells. Anticancer Res. 2018; 38(10):5679-5684.
- Chan EWC, Soo OY, Tan YH, Wong SK, Chan HT. Nobiletin and tangeretin (citrus polymethoxyflavones): An overview on their chemistry, pharmacology and cytotoxic activities against breast cancer. J Chin Pharm Sci. 2020; 29(7):443-454.
- Semwal RB, Semwal DK, Combrinck S, Trill J, Gibbons S, Viljoen A. Acacetin A simple flavone exhibiting diverse pharmacological activities. Phytochem Lett. 2019; 32:56-65.
- 13. Singh S, Gupta P, Meena A, Luqman S. Acacetin, a flavone with diverse therapeutic potential in cancer, inflammation, infections and other metabolic disorders. Food Chem Toxicol. 2020; 145:111708.
- Chan EWC, Ng YK, Tan CY, Alessandro L, Wong SK, Chan HT. Diosmetin and tamarixetin (methylated flavonoids): A review on their chemistry, sources, pharmacology, and anticancer properties. J Appl Pharm Sci. 2021; 11(3):22-28.
- Kwon HS, Ha TJ, Hwang SW, Jin YM, Nam SH, Park KH, Yang MS. Cytotoxic flavonoids from the whole plants of *Chrysanthemum zawadskii* Herbich var. *latilobum* Kitamura. J Life Sci. 2006; 16(5):746-749.
- González-Trujano ME, Ventura-Martínez R, Chávez M, Díaz-Reval I, Pellicer F. Spasmolytic and antinociceptive activities of ursolic acid and acacetin identified in *Agastache mexicana*. Planta Med. 2012; 78(8):793-796.
- Rauf A, Khan R, Khan H, Ullah B, Pervez S. Antipyretic and antinociceptive potential of extract/fractions of *Potentilla evestita* and its isolated compound, acacetin. BMC Compl Altern Med. 2014; 14(1):448.
- Zou GA, Guo D, Zhao HQ, Aisa HA. Bioactive constituents of *Ziziphora clinopodioides*. Chem Nat Compd. 2015; 51(5):961-963.
- 19. Veitch NC, Elliott PC, Kite GC, Lewis GP. Flavonoid glycosides of the black locust tree, *Robinia pseudoacacia* (Leguminosae). Phytochem. 2010; 71(4):479-486.
- Al-Hazimi HM and Basha RM. Phenolic compounds from various *Artemisia* species. J Chem Soc Pak. 1991; 13(4):277-289.

- Liou CJ, Wu SJ, Chen LC, Yeh KW, Chen CY, Huang WC. Acacetin from traditionally used *Saussurea involucrata* Kar. et Kir. suppressed adipogenesis in 3T3-L1 adipocytes and attenuated lipid accumulation in obese mice. Front Pharmacol. 2017; 8:589.
- 22. Suksamrarn A, Chotipong A, Suavansri T, Boongird S, Timsuksai P, Vimuttipong S, Chuaynugul A. Antimycobacterial activity and cytotoxicity of flavonoids from the flowers of *Chromolaena odorata*. Arch Pharm Res. 2004; 27(5):507-511.
- 23. Lin LZ and Harnly JM. Identification of the phenolic components of chrysanthemum flower (*Chrysanthemum morifolium* Ramat). Food Chem. 2010; 120(1):319-326.
- Kim EO, Oh JH, Lee SK, Lee JY, Choi SW. Antioxidant properties and quantification of phenolic compounds from safflower (*Carthamus tinctorius* L.) seeds. Food Sci Biotechnol. 2007; 16(1):71-77.
- Yin J, Ma Y, Liang C, Gao J, Wang H, Zhang L. A systematic study of the metabolites of dietary acacetin *in* vivo and *in vitro* based on UHPLC-Q-TOF-MS/MS analysis. J Agric Food Chem. 2019; 67(19):5530-5543.
- Wu X, Zhang S, Li X, Zhang F, Fan Y, Liu Q, Wan X, Lin T. Postharvest UV-B radiation increases enzyme activity, polysaccharide and secondary metabolites in honeysuckle (*Lonicera japonica* Thunb.). Ind Crops Prod. 2021; 171:113907.
- Bramati L, Minoggio M, Gardana C, Simonetti P, Mauri P, Pietta P. Quantitative characterization of flavonoid compounds in rooibos tea (*Aspalathus linearis*) by LC– UV/DAD. J Agric Food Chem. 2002; 50(20):5513-5519.
- 28. Khan AU and Gilani AH. Selective bronchodilatory effect of rooibos tea (*Aspalathus linearis*) and its flavonoid, chrysoeriol. Eur J Nutr. 2006; 45(8):463.
- Goławska S, Łukasik I, Kapusta T, Janda B. Analysis of flavonoids content in alfalfa. Ecol Chem Eng A. 2010; 17(2-3):261-267.
- Tai BH, Cuong NM, Huong TT, Choi EM, Kim JA, Kim YH. Chrysoeriol isolated from the leaves of *Eurya ciliata* stimulates proliferation and differentiation of osteoblastic MC3T3-E1 cells. J Asian Nat Prod Res. 2009; 11(9):817-823.
- Kim YH, Lee YS, Choi EM. Chrysoeriol isolated from Eurya cilliata leaves protects MC3T3-E1 cells against hydrogen peroxide-induced inhibition of osteoblastic differentiation. J Appl Toxicol. 2010; 30(7):666-673.
- Chen SM, Li CH, Zhu XR, Deng YM, Sun W, Wang LS, Chen FD, Zhang Z. The identification of flavonoids and the expression of genes of anthocyanin biosynthesis in the chrysanthemum flowers. Biol Plant. 2012; 56(3):458-464.
- Hsu YL, Kuo PL, Lin CC. Acacetin inhibits the proliferation of Hep G2 by blocking cell cycle progression and inducing apoptosis. Biochem Pharmacol. 2004; 67(5):823-829.
- Shim HY, Park JH, Paik HD, Nah SY, Kim DS, Han YS. Acacetin-induced apoptosis of human breast cancer MCF-7 cells involves caspase cascade, mitochondria-mediated death signalling and SAPK/JNK1/2-c-Jun activation. Mol Cells. 2007; 24(1):95-104.
- Watanabe K, Kanno SI, Tomizawa A, Yomogida S, Ishikawa M. Acacetin induces apoptosis in human T cell leukaemia Jurkat cells *via* activation of a caspase cascade. Oncol Rep. 2012; 27(1):204-209.
- Kim CD, Cha JD, Li S, Cha IH. The mechanism of acacetin-induced apoptosis on oral squamous cell carcinoma. Arch Oral Biol. 2015; 60(9):1283-1298.
- Hsu YL, Kuo PL, Liu CF, Lin CC. Acacetin-induced cell cycle arrest and apoptosis in human non-small cell lung cancer A549 cells. Cancer Lett. 2004; 212(1):53-60.
- Pan MH, Lai CS, Hsu PC, Wang YJ. Acacetin induces apoptosis in human gastric carcinoma cells accompanied by activation of caspase cascades and production of

reactive oxygen species. J Agric Food Chem. 2005; 53(3):620-630.

- 39. Singh RP, Agrawal P, Yim D, Agarwal C, Agarwal R. Acacetin inhibits cell growth and cell cycle progression, and induces apoptosis in human prostate cancer cells: Structure-activity relationship with linarin and linarin acetate. Carcinogenesis. 2005; 26(4):845-854.
- Fong Y, Shen KH, Chiang TA, Shih YW. Acacetin inhibits TPA-induced MMP-2 and u-PA expressions of human lung cancer cells through inactivating JNK signalling pathway and reducing binding activities of NFκB and AP-1. J Food Sci. 2010; 75(1):30-38.
- 41. Shen KH, Hung SH, Yin LT, Huang CS, Chao CH, Liu CL, Shih YW. Acacetin, a flavonoid, inhibits the invasion and migration of human prostate cancer DU145 cells *via* inactivation of the p38 MAPK signalling pathway. Mol Cell Biochem. 2010; 333:279-291.
- 42. Chien ST, Lin SS, Wang CK, Lee YB, Chen KS, Fong Y, Shih YW. Acacetin inhibits the invasion and migration of human non-small cell lung cancer A549 cells by suppressing the p38α MAPK signalling pathway. Mol Cell Biochem. 2011; 350:135-148.
- Kim HR, Park CG, Jung JY. Acacetin (5,7-dihydroxy-4'methoxyflavone) exhibits *in vitro* and *in vivo* anticancer activity through the suppression of NF-κB/Akt signalling in prostate cancer cells. Int J Mol Med. 2014; 33(2):317-324.
- 44. Salimi A, Roudkenar MH, Sadeghi L, Mohseni A, Seydi E, Pirahmadi N, Pourahmad J. Selective anticancer activity of acacetin against chronic lymphocytic leukaemia using both *in vivo* and *in vitro* methods: Key role of oxidative stress and cancerous mitochondria. Nutr Cancer. 2016; 68(8):1404-1416.
- Sun F, Li D, Wang C, Peng C, Zheng H, Wang X. Acacetin-induced cell apoptosis in head and neck squamous cell carcinoma cells: Evidence for the role of muscarinic M3 receptor. Phytother Res. 2019; 33(5):1551-1561.
- 46. Kang KR, Kim JS, Kim TH, Seo JY, Park JH, Lim JW, Yu SK, Kim HJ, Shin SH, Park BR, Kim CS. Inhibition of cell growth and induction of apoptosis by acacetin in FaDu human pharyngeal carcinoma cells. Int J Oral Biol. 2020; 45(3):107-114.
- 47. Prasad N, Sharma JR, Yadav UC. Induction of growth cessation by acacetin *via* β -catenin pathway and apoptosis by apoptosis inducing factor activation in colorectal carcinoma cells. Mol Biol Rep. 2020; 47(2):987-1001.
- Shendge AK, Chaudhuri D, Mandal N. The natural flavones, acacetin and apigenin, induce Cdk-cyclin mediated G2/M phase arrest and trigger ROS-mediated apoptosis in glioblastoma cells. Mol Biol Rep. 2021; 48(1):539-549.
- 49. Yang Y, Zhou X, Xiao M, Hong Z, Gong Q, Jiang L, Zhou J. Discovery of chrysoeriol, a PI3K-AKT-mTOR pathway inhibitor with potent antitumor activity against human multiple myeloma cells *in vitro*. J Huazhong Univ Sci Technol [Med Sci]. 2010; 30(6):734-740.
- Ninomiya M, Nishida K, Tanaka K, Watanabe K, Koketsu M. Structure-activity relationship studies of 5,7dihydroxyflavones as naturally occurring inhibitors of cell proliferation in human leukaemia HL-60 cells. J Nat Med. 2013; 67(3):460-467.
- 51. Wei W, He J, Ruan H, Wang Y. *In vitro* and *in vivo* cytotoxic effects of chrysoeriol in human lung carcinoma are facilitated through activation of autophagy, sub-G1/G0 cell cycle arrest, cell migration and invasion inhibition and modulation of MAPK/ERK signalling pathway. J BUON. 2019; 24(3):936-942.
- Manthey JA, Guthrie N. Antiproliferative activities of citrus flavonoids against six human cancer cell lines. J Agric Food Chem. 2002; 50(21):5837-5843.

- Amrutha K, Nanjan P, Shaji SK, Sunilkumar D, Subhalakshmi K, Rajakrishna L, Banerji A. Discovery of lesser-known flavones as inhibitors of NF-κB signalling in MDA-MB-231 breast cancer cells – A SAR study. Bioorg Med Chem Lett. 2014; 24(19):4735-4742.
- Zhang Y, Li Z, Min Q, Palida A, Zhang Y, Tang R, Chen L, Li H. 8-Chrysoeriol, as a potential BCL- inhibitor, triggers apoptosis of SW1990 pancreatic cancer cells. Bioorg Chem. 2018; 77:478-484.
- 55. Takemura H, Nagayoshi H, Matsuda T, Sakakibara H, Morita M, Matsui A, Ohura T, Shimoi K. Inhibitory effects of chrysoeriol on DNA adduct formation with benzo[α]pyrene in MCF-7 breast cancer cells. Toxicology. 2010; 274(1-3):42-48.
- Takemura H, Uchiyama H, Ohura T, Sakakibara H, Kuruto R, Amagai T, Shimoi K. A methoxyflavonoid, chrysoeriol, selectively inhibits the formation of a carcinogenic estrogen metabolite in MCF-7 breast cancer cells. J Steroid Biochem Mol Biol. 2010; 118(1-2):70-76.
- 57. Tan KW, Li Y, Paxton JW, Birch NP, Scheepens A. Identification of novel dietary phytochemicals inhibiting the efflux transporter breast cancer resistance protein (BCRP/ABCG2). Food Chem. 2013; 138(4):2267-2274.
- Min DY, Jung E, Ahn SS, Lee YH, Lim Y, Shin SY. Chrysoeriol prevents TNFα-induced Cyp19 gene expression *via* Egr-1 downregulation in MCF-7 breast cancer cells. Int J Mol Sci. 2020; 21(20):7523.
- Li S, Lv Q, Sun X, Tang T, Deng X, Yin Y, Li L. Acacetin inhibits *Streptococcus pneumoniae* virulence by targeting pneumolysin. J Pharm Pharmacol. 2020; 72(8):1092-1100.
- Hayashi K, Hayashi T, Arisawa M, Morita N. Antiviral agents of plant origin: Antiherpetic activity of acacetin. Antiviral Chem Chemother. 1993; 4(1):49-53.
- Kwon EB, Kang MJ, Ryu HW, Lee S, Lee JW, Lee MK, Lee HS, Lee SU, Oh SR, Kim MO. Acacetin enhances glucose uptake through insulin-independent GLUT4 translocation in L6 myotubes. Phytomed. 2020; 68:153178.
- 62. Ha SK, Moon E, Lee P, Ryu JH, Oh MS, Kim SY. Acacetin attenuates neuroinflammation *via* regulation the response to LPS stimuli *in vitro* and *in vivo*. Neurochem Res. 2012; 37(7):1560-1567.
- 63. Kim HG, Ju MS, Ha SK, Lee H, Lee H, Kim SY, Oh MS. Acacetin protects dopaminergic cells against 1-methyl-4phenyl-1,2,3,6-tetrahydropyridine-induced neuroinflammation *in vitro* and *in vivo*. Biol Pharm Bull. 2012; 35(8):1287-1294.
- Chen WP, Yang ZG, Hu PF, Bao JP, Wu LD. Acacetin inhibits expression of matrix metalloproteinases *via* a MAPK-dependent mechanism in fibroblast-like synoviocytes. J Cell Mol Med. 2015; 19(8):1910-1915.
- Asthana J, Mishra BN, Pandey R. Acacetin promotes healthy aging by altering stress response in *Caenorhabditis elegans*. Free Radic Res. 2016; 50(8):861-874.
- 66. Wang X, Perumalsamy H, Kwon HW, Na YE, Ahn YJ. Effects and possible mechanisms of action of acacetin on the behaviour and eye morphology of *Drosophila* models of Alzheimer's disease. Sci Rep. 2015; 5:16127.
- Carballo-Villalobos AI, González-Trujano ME, López-Muñoz FJ. Evidence of mechanism of action of antiinflammatory/antinociceptive activities of acacetin. Eur J Pain. 2014; 18(3):396-405.
- Shahidi F, Ramakrishnan VV, Oh WY. Bioavailability and metabolism of food bioactives and their health effects: A review. J Food Bioact. 2019; 8:6-41.
- Barreca D, Mandalari G, Calderaro A, Smeriglio A, Trombetta D, Felice MR, Gattuso G. *Citrus* flavones: An update on sources, biological functions, and health promoting properties. Plants. 2020; 9(3):288.

- Ramirez G, Zamilpa A, Zavala M, Perez J, Morales D, Tortoriello J. Chrysoeriol and other polyphenols from *Tecoma stans* with lipase inhibitory activity. J Ethnopharmacol. 2016; 185:1-8.
- Krishnan B, Ganesan AR, Balasubramani R, Nguyen DD, Chang SW, Wang S, Xiao J, Balasubramanian B. Chrysoeriol ameliorates hyperglycaemia by regulating the carbohydrate metabolic enzymes in streptozotocin-induced diabetic rats. Food Sci Hum Wellness. 2020; 9(4):236-354.
- 72. Bashyal P, Parajuli P, Pandey RP, Sohng JK. Microbial biosynthesis of antibacterial chrysoeriol in recombinant *Escherichia coli* and bioactivity assessment. Catalysts. 2019; 9(2):112.
- Choi DY, Lee JY, Kim MR, Woo ER, Kim YG, Kang KW. Chrysoeriol potently inhibits the induction of nitric oxide synthase by blocking AP-1 activation. J Biomed Sci. 2005; 12(6):949-959.
- Wu JY, Chen YJ, Bai L, Liu YX, Fu XQ, Zhu PL, Li JK, Chou JY, Yin CL, Wang YP, Bai JX. Chrysoeriol ameliorates TPA-induced acute skin inflammation in mice

and inhibits NF- κ B and STAT3 pathways. Phytomed. 2020; 68:153173.

- 75. Yoon HS and Park CM. Chrysoeriol ameliorates COX-2 expression through NF-κB, AP-1 and MAPK regulation *via* the TLR4/MyD88 signalling pathway in LPS-stimulated murine macrophages. Exper Ther Med. 2021; 22(1):1-6.
- Limboonreung T, Tuchinda P, Chongthammakun S. Chrysoeriol mediates mitochondrial protection *via* PI3K/Akt pathway in MPP⁺ treated SH-SY5Y cells. Neurosci Lett. 2020; 714:134545.
- 77. Yokozawa T, Chen CP, Dong E, Tanaka T, Nonaka GI, Nishioka I. Study on the inhibitory effect of tannins and flavonoids against the 1,1-diphenyl-2-picrylhydrazyl radical. Biochem Pharmacol. 1998; 56(2):213-222.
- Lai S, Chen JN, Huang HW, Zhang XY, Jiang HL, Li W, Wang PL, Wang J, Liu FN. Structure activity relationships of chrysoeriol and analogs as dual c-Met and VEGFR2 tyrosine kinase inhibitors. Oncol Rep. 2018; 40(3):1650-1656.